

Design and Synthesis of 2-(3-Benzo[*b*]thienyl)-6,7-methylenedioxyquinolin-4-one Analogues as Potent Antitumor Agents that Inhibit Tubulin Assembly

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As part of our continuing investigation of azo-flavonoid derivatives as potential anticancer drug candidates, a series of 2-aryl-6,7-methylenedioxyquinolin-4-one analogues was designed and synthesized. The design combined structural features of 2-(2-fluorophenyl)-6,7-methylenedioxyquinolin-4-one (**CHM-1**), a previously discovered compound with potent *in vivo* antitumor activity, and 2-arylquinolin-4-ones, identified by CoMFA models. The newly synthesized analogues were evaluated for cytotoxicity against seven human cancer cell lines, and structure–activity relationship (SAR) correlations were established. Analogues **1**, **37**, and **39** showed potent cytotoxicity against different cancer cell lines. Compound **1** demonstrated selective cytotoxicity against Hep 3B (hepatoma) cells. Compound **37** was cytotoxic against HL-60 (leukemia), HCT-116 (colon cancer), Hep 3B (hepatoma), and SK-MEL-5 (melanoma) cells. Compound **39** exhibited broad cytotoxicity against all seven cancer cell lines, with IC₅₀ values between 0.07 and 0.19 μM. Results from mechanism of action studies revealed that these new quinolone derivatives function as antitubulin agents.

Introduction

Microtubules of eukaryotic cells are known to play an important role in mitosis.¹ Therefore, compounds that target microtubules have long been investigated as anticancer drugs. Typically, such compounds arrest cells, at least transiently, in the mitotic phase of the cell cycle. Currently, two groups of antimitotic agents are used clinically for cancer treatment. Vinca alkaloids and estramustine inhibit tubulin polymerization into microtubules, while taxoids and epothilones stabilize microtubules and consequently interfere with their normal dynamic instability.^{2,3} Colchicine is another well-known antimitotic agent used for treating gout and inflammatory diseases. However, because of its high toxicity, colchicine is not used in cancer therapy. Combretastatin A-4, with a modified colchicine-like structure, exhibits a lower toxicity profile while it still targets the same binding site as colchicine.^{4,5} The phosphate prodrug of combretastatin A-4 (CA-4) is currently in phase III clinical trials as an anticancer agent.^{6,7} Although several antimitotic agents are available, due to the structural complexity of vinca alkaloids and taxoids, there is still a need to identify novel anticancer drugs that target microtubules but have simplified structures because such compounds can be readily prepared in a more cost-effective way.

In recent years, we have designed and synthesized three series of azo-flavonoids as new classes of antimitotic agents:

2-arylquinolin-4-ones,^{8–15} 2-arylnaphthyridin-4-ones,^{16–18} and 2-arylquinazolin-4-ones^{19,20} (Figure 1). Some of these compounds, especially analogues belonging to the 2-arylnaphthyridin-4-one family, demonstrated significant antitumor and antitubulin activities. In general, a good correlation was found between the cytotoxicity of these analogues and their inhibitory effects on tubulin polymerization. Additional mechanism of action studies revealed that the most active of these compounds are also potent inhibitors of colchicine binding to tubulin. Further modification of the 2-phenylquinolin-4-one series was directed by computer modeling studies combined with SAR results from previously synthesized compounds. In the current study, we report the design and syntheses of novel 2-arylquinolin-4-one analogues, which showed potent and selective inhibitory activities toward different human cancer cell lines.

Design. Both conventional comparative molecular field analysis (CoMFA) and *g*² GRS CoMFA were used to identify structural requirements that may be essential for increasing the binding affinity of 2-phenylquinolin-4-one and 2-phenylnaphthyridin-4-one analogues for the colchicine site of tubulin. The training set of the models contained 51 compounds, and the cross-validated *R*² (*q*²) values for conventional CoMFA and *g*² GRS CoMFA were 0.637 and 0.692, respectively. These QSAR models predicted that a larger heterocyclic aromatic ring at the C-2 position should enhance activity. The predictive power of the models was validated by the prediction for a test set of 53 compounds with known antitubulin potencies, and the predictive *R*² values were 0.546 and 0.426, respectively. On the basis of this study, new analogues were designed in both compound

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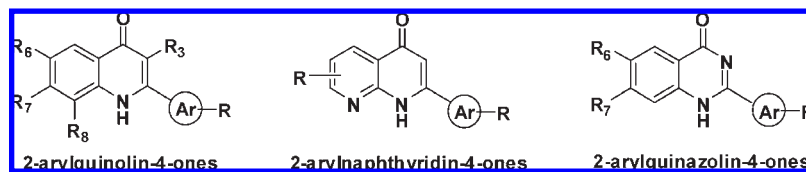


Figure 1. Three series of structural skeletons of azo-flavonoids: 2-arylquinolin-4-ones, 2-arylnaphthyridin-4-ones, and 2-arylquinazolin-4-ones.

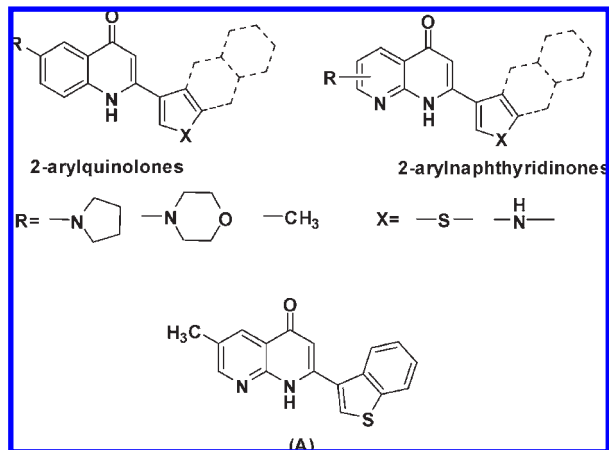


Figure 2. Proposed models and newly designed compounds. Conventional comparative molecular field analysis (CoMFA) and g^2 GRS CoMFA are two computational programs performed to identify the essential structure requirements for increasing affinity at the colchicine site of tubulin. On the basis of the results of the QSAR study, we proposed two concept models. As the proof of concept, analogue **A** was synthesized and tested against cancer cell lines and for binding to tubulin.

series (Figure 2).²¹ As a proof of concept, the newly designed arylnaphthyridine analogue 2-(3-benzo[*b*]thienyl)naphthyridin-4-one (**A**), which has a benzothienyl ring at the C-2 position of the naphthyridine, exhibited potent cytotoxicity in the low micromolar to nanomolar concentration range. Further, a mechanism of action study revealed that the compound also inhibited tubulin polymerization with an IC_{50} value of 0.37 μ M. These results verified the success of our modeling design.¹⁸ Thus, the same modifications were carried out at the C-2 position of the 2-arylquinolin-4-one series in the present study.

In addition, when we reanalyzed results from our prior *in vivo* antitumor study of selected 2-phenylquinolin-4-one compounds, we found that analogues with a methylenedioxy functional group at the C-6 and -7 positions showed superior antitumor and safety profiles compared with C-6 monomodified compounds. For instance, 2-(3-methoxyphenyl)-6-pyrrolinylquinolin-4-one (**B**, Figure 3) exhibited only moderate antitumor activity at the maximum tolerated dose (MTD, 25 mg/kg, ip, once weekly for three courses) against the OVCAR-3 xenograft model in nude mice. In contrast, 2-(2-fluorophenyl)-6,7-methylenedioxyquinolin-4-one (**CHM-1**, Figure 3) extended the life span of tumor-bearing mice by 124%, 133%, and 79%, at dosages of 200, 134, and 79 mg/kg (ip, once weekly for three courses), respectively. Additionally, the maximum tolerated dose of **CHM-1** was not reached at the highest tested dosage of 200 mg/kg.²² Currently, the phosphate prodrug of **CHM-1** (**CHM-1-P-Na**, Figure 4) is under preclinical study.^{22,23} Therefore, the methylenedioxy ring of **CHM-1** was also incorporated into our newly designed 2-arylquinolin-4-one series, which led

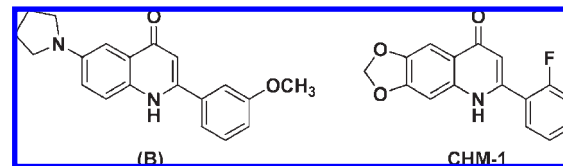


Figure 3. Structures of compound **B** and **CHM-1**.

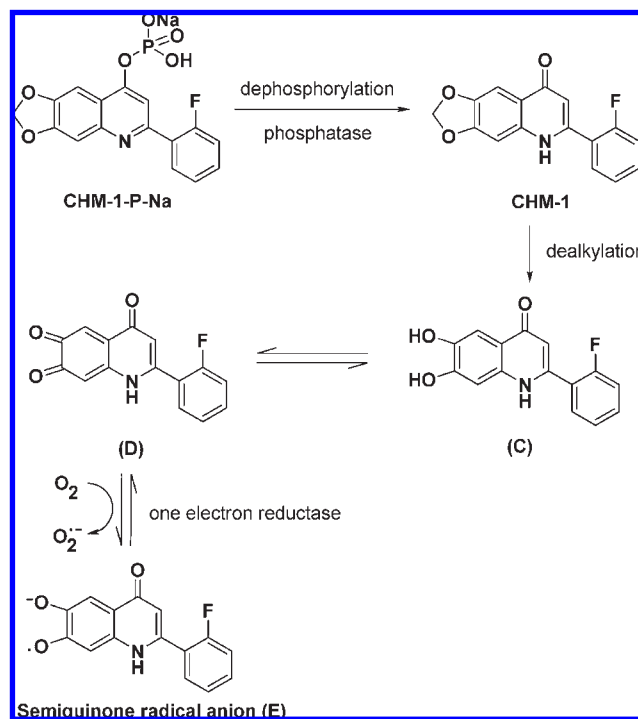
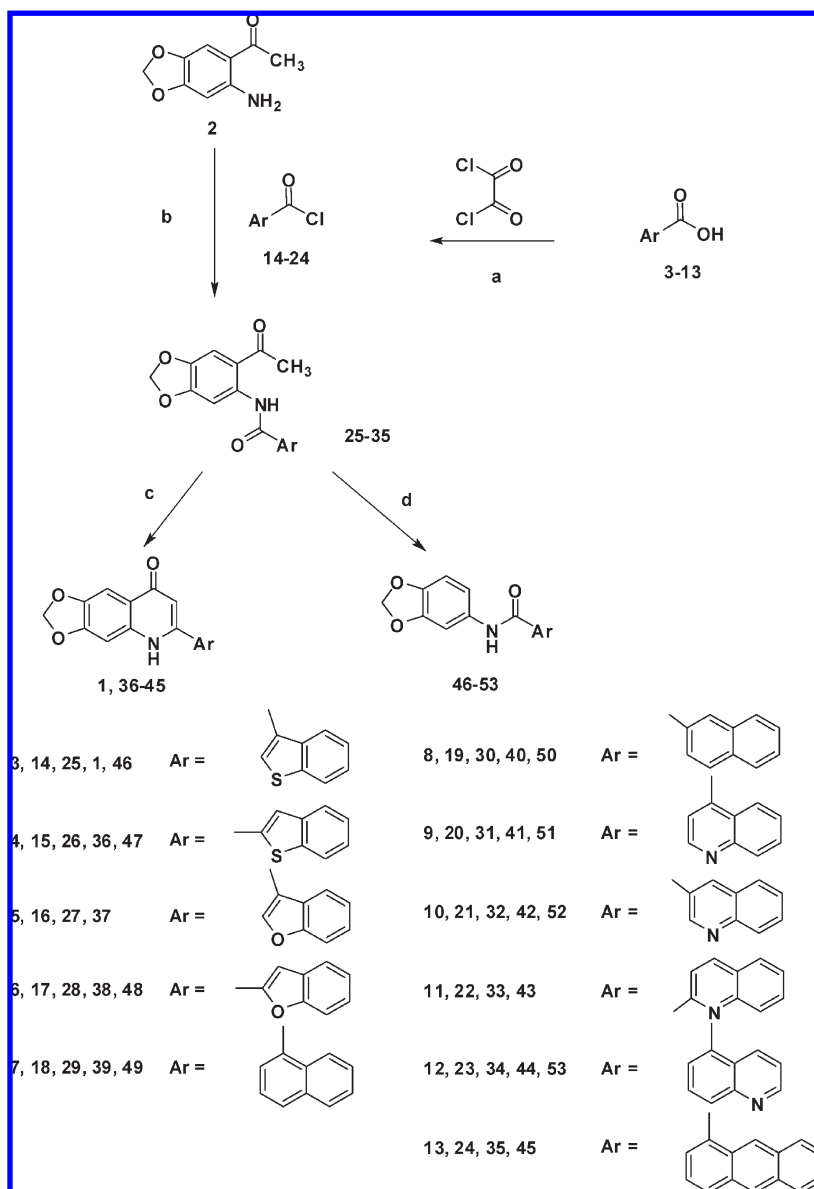


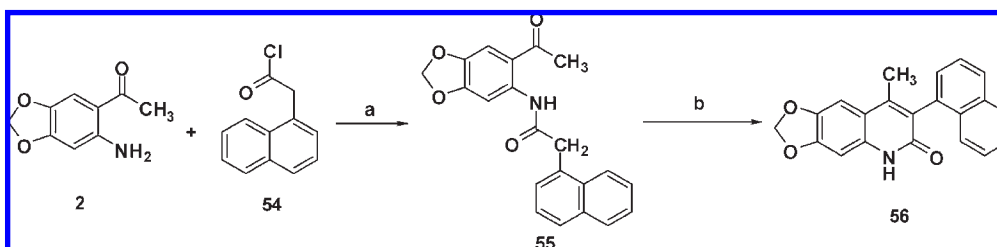
Figure 4. Metabolic pathway of **CHM-1-P-Na**.

to the syntheses of 2-aryl-6,7-methylenedioxyquinolin-4-one analogues (**1**, **36–45**). Compound **56**, a ring isomer of analogue **39**, was also synthesized because ring isomerization is a common approach in structure–activity relationship studies of a compound series.

Chemistry. The synthetic procedure for target compounds (**1**, **36–45**) is illustrated in Scheme 1. The starting arylcarboxylic acids (**3–13**) were first treated with oxalyl chloride to form the corresponding acid chlorides (**14–24**), which were then reacted with 2-amino-4,5-methylenedioxyacetophenone (**2**) to give the corresponding amides (**25–35**). The intermediates (**25**, **27–34**) were subjected to a cyclization reaction in *t*-BuOH, in the presence of *t*-BuOK, to afford the corresponding 2-arylquinolin-4-one analogues (**1**, **37–44**). Cyclization was also carried out with 1,4-dioxane as solvent, in the presence of NaOH, which yielded the target products (**1**, **36**, **39**, and **45**) in yields of 40–74%. When the starting compounds (**25–26**, **28–32**, and **34**) were treated with *t*-BuOK in toluene at 90 °C for 72 h, the

Scheme 1^a

^a Reagents and conditions: (a) toluene/DMF, 22–25 °C; (b) toluene/triethylamine, 22–25 °C; (c) *t*-BuOK/*t*-BuOH or NaOH/1,4-dioxane, reflux; (d) *t*-BuOK/toluene, 90 °C, 72 h.

Scheme 2^a

^a Reagents and conditions: (a) toluene/triethylamine, 22–25 °C; (b) NaOH/1,4-dioxane, reflux.

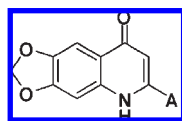
corresponding deacetylated products **46–53** were obtained unexpectedly.

Scheme 2 depicts the synthesis of compound **56**. The starting material **2** was first treated with 1-naphthylmethyl chloride (**54**) to give the corresponding amide (**55**). Compound **55** was then subjected to a cyclization reaction in 1,4-dioxane, in the presence of NaOH, which

yielded 4-methyl-6,7-methylenedioxy-3-(1-naphthyl)quinolin-2-one (**56**) as the final product.

Results and Discussion

The newly synthesized 2-aryl-6,7-methylenedioxyquinolin-4-ones (**1, 36–45**) and the ring isomer **56** were assayed for

Table 1. IC₅₀ (μM) Values from *In Vitro* Cytotoxicity Testing of **1**, **36**–**45**, and **56**

Compound	Ar	HL-60	HCT-116	A549	Hep 3B	KB	Kb-VIN	DU145
1		0.17	0.14	>0.25	0.06	>0.25	>0.25	>0.25
36		>1.00	>1.00	>1.00	>1.00	>1.00	>1.00	>1.00
37		0.03	0.05	2.98	0.09	1.05	0.59	1.87
38		>1.00	>1.00	>1.00	>1.00	>1.00	>1.00	>1.00
39		0.07	0.07	0.13	0.07	0.13	0.19	0.13
40		0.50	>1.00	>1.00	>1.00	>1.00	>1.00	>1.00
41		>5.00	>5.00	>5.00	>5.00	>5.00	>5.00	>5.00
42		>0.10	>0.10	>0.10	>0.10	>0.10	>0.10	>0.10
43		>5.00	>5.00	>5.00	>5.00	>5.00	>5.00	>5.00
44		>10.00	>10.00	>10.00	>10.00	>10.00	>10.00	>10.00
45		0.53	1.60	NA*	2.20	NA	NA	NA
56		>10.00	>10.00	NA	4.30	NA	NA	NA

* Not assayed

in vitro cytotoxicity against seven human cancer cell lines, including HL-60 (leukemia), HCT-116 (colon cancer), A549 (nonsmall cell lung carcinoma), Hep 3B (hepatoma), KB (epidermoid carcinoma of the nasopharynx), KB-VIN (p-glycoprotein-expressing epidermoid carcinoma of the nasopharynx), and DU145 (prostate cancer). The cytotoxicity results are summarized in Table 1. Compounds **46**–**53** were tested against the HL-60, HCT-116, H226, A549, Hep G2, and A498 cancer cell lines; however, none of the compounds showed significant activity (IC₅₀ values were > 10 μM).

Compound **1** exhibited significant cytotoxicity against Hep 3B cells, with an IC₅₀ value of 0.06 μM, and moderate activity against HL-60 and HCT-116 cells, with IC₅₀ values of 0.17 and 0.14 μM, respectively. This result, along with the prior cytotoxicity data obtained with compound **A**, confirmed the prediction generated by conventional and g² GRS CoMFA that a larger heterocyclic aromatic ring should be preferred at the C-2 position. In comparison, replacement of the 3-benzofuran group (**1**) with a 2-benzofuran group (**36**)

resulted in significantly reduced cytotoxicity, demonstrating the importance of the exact linkage of the aromatic ring moiety.

Bioisosteric replacement of 3-benzofuran (**1**) with 3-benzofuran (**37**) led to increased potency against two cancer cell lines. Specifically, compared with **1**, the cytotoxic activity of **37** increased 5-fold against HL-60 (IC₅₀ 0.03 μM) and 3-fold against HCT-116 (IC₅₀ 0.05 μM) cells. The two compounds had similar potency against Hep 3B cells. Most importantly, **37** retained potency (IC₅₀: 0.59 μM) against KB-VIN cells, a vincristine-resistant epidermoid carcinoma of the nasopharynx cell line, compared with KB cells (IC₅₀ 1.05 μM). As with **1** and **36**, moving the attachment of the benzofuran moiety from 3' in **37** to 2' in **38** was detrimental to cytotoxic activity.

Compound **39**, with a 1-naphthyl group rather than a 3-benzofuran group (**1**) at the C-2 position, exhibited potent cytotoxicity toward all seven cancer cell lines, with IC₅₀ values ranging from 0.07 to 0.19 μM. It should be noted that although **39** showed similar activity toward the HL-60, HCT-116, and

Table 2. Anti-Tubulin Data for CA-4, **1**, and **37-45**

compd	inhibition of tubulin assembly ^a IC ₅₀ (μM) ± SD	inhibition of colchicine binding ^b % inhibition at tested concentration	
		1 μM	5 μM
CA-4	1.2 ± 0.2	91	99
1	0.76 ± 0.06	25	61
37	0.58 ± 0.02	39	51
38	> 4		
39	0.64 ± 0.07	33	67
40	> 40		
41	> 40		
42	> 4		
43	> 40		
44	27 ± 3		
45	> 40		

^a Assembly assay contained 10 μM tubulin. ^b Colchicine binding assay contained 1 μM tubulin and 5 μM [³H]colchicine.

Hep 3B cancer cell lines as compared with **1** and **37**, it showed significantly increased cytotoxicity toward A549, KB, KB-VIN, and DU145 cells. Compound **40**, the ring positional isomer of **39**, was much less cytotoxic, as was the case with compounds **36** and **38**.

Surprisingly, cytotoxicity was reduced significantly when the C-2 aryl moiety was a quinoline group, no matter how it was attached (**41-44**). In addition, expanding the naphthalene ring of **39** to an anthracene (**45**) lowered the cytotoxicity remarkably. Thus, we speculate that the size of the C-2 substituted aromatic ring plays an important role in antitumor activity and that the binding pocket for this portion of the drug molecule is quite small.

Compound **56** was designed as a ring isomer of **39**. The *in vitro* bioassay data showed that moving the aromatic ring to C-3 and the carbonyl to C-2 to give a cyclic amide completely abolished cytotoxicity of the compound. Nuncyclic amide analogues **46-53** also exhibited poor *in vitro* activity.

In addition to the *in vitro* cytotoxicity study, we also performed initial mechanism of action studies with the newly synthesized 2-aryl-6,7-methylenedioxyquinolin-4-one analogues (**1**, **37-45**). As noted above, 2-arylquinolin-4-ones are azo-flavonoids, which were shown to inhibit tubulin assembly and the binding of colchicine to tubulin. Therefore, compounds **1** and **37-45** were tested for their *in vitro* activities in these assays in comparison with combretastatin A-4 (CA-4). The results are summarized in Table 2. The data showed that **1**, **37**, and **39**, the three compounds with the greatest *in vitro* cytotoxicity, were potent inhibitors of tubulin assembly, with IC₅₀ values of 0.76, 0.58, and 0.64 μM, respectively. Although all three compounds were better assembly inhibitors than CA-4 (IC₅₀ 1.2 μM), they were less effective than CA-4 in inhibiting colchicine binding to tubulin.

Furthermore, results from a pharmacokinetic study in a mice model revealed that the 6,7-methylenedioxy moiety of CHM-1-P-Na is metabolized to an *ortho*-quinone (**D**, Figure 4, unpublished data). It is known that the *para*-quinone moiety in mitomycin C can be subjected to one-electron reduction by NADPH-cytochrome C (P450) reductase to form the corresponding semiquinone radical anion.²⁴ We postulate that similar reduction of the *ortho*-quinone moiety of **D** may take place to form the radical anion product **E**, which may be further metabolized or broken down into more cytotoxic metabolites in hypoxic cells. Because severe hypoxia is a common property of locally advanced solid tumors, this postulate may explain our finding that 6,7-methylenedioxyquinoline analogues (e.g., CHM-1) showed enhanced *in vivo* activity profiles compared with 6-monosubstituted analogues.

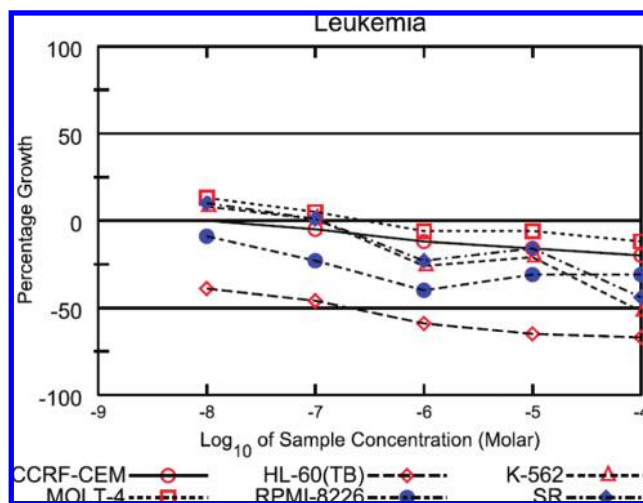


Figure 5. Dose-response curves of **37** against different leukemia cell lines.

Among the active analogues, **37**, which showed potent cytotoxicity and reasonable solubility, was chosen for submission to the National Cancer Institute (NCI, USA) for further screening against 60 human tumor cell lines. In preliminary screening, **37** showed potent selective cytotoxicity against many leukemia cell lines (Figure 5). In addition, it also significantly inhibited the growth of several colon cancer (HCC-2998 and KM-12), CNS cancer (SF-539 and SNB-75), melanoma (SK-MEL-5), and ovarian cancer (IGROV1 and OVCAR-3) cell lines. On the basis of these results, **37** appears to be an attractive candidate for further development as a potential anticancer agent in the clinic.

In conclusion, a series of 2-aryl-6,7-methylenedioxyquinolin-4-one analogues (**1**, **36-45**) were designed, synthesized, and evaluated for *in vitro* cytotoxicity. This design combined two structural features: larger heterocyclic aromatic rings at the C-2 position as predicted to be favorable by CoMFA models and a 6,7-methylenedioxy moiety rather than C-6 monosubstituted analogue based on the improved cytotoxicity of CHM-1 and the metabolic pathway of CHM-1-P-Na. In our studies, **1** showed selective cytotoxicity against Hep 3B cells, **37** was active against HL-60, HCT-116, Hep 3B, and KB-VIN cells, and **39** had potent cytotoxicity against all seven cancer cell lines, with IC₅₀ values between 0.07 and 0.19 μM. A mechanism of action study demonstrated that **1**, **37**, and **39** also function as antitubulin agents. Compound **37** was further selected for evaluation against 60 human cancer cell lines and

was active against several types of cancer cells. The significant *in vitro* cytotoxicity of **37** and **39** suggested that they can be further developed as anticancer drugs.

Experimental Section

Chemistry. General Experimental Procedures. Reagents and solvents were obtained commercially and used without further purification. Reactions were monitored by thin-layer chromatography, using Merck plates with fluorescent indicator (TLC Silica Gel 60 F₂₅₄). Flash column chromatography was performed on silica gel (Merck Silica Gel 60, 40–63 μm) using a mixture of CH₂Cl₂ and EtOH as eluant. Melting points were determined on a Yanaco MP-500D melting point apparatus and were uncorrected. IR spectra were recorded on a Shimadzu IRPrestige-21 spectrophotometer as KBr pellets. ¹H NMR spectra were obtained on a Bruker NMR AV 400 spectrometer in DMSO. The following abbreviations are used: s, singlet; d, doublet; t, triplet; m, multiplet; dd, double doublet; td, triple doublet; ddd, double double doublet. EI-MS spectra were measured with an HP 5995 GC-MS instrument. ESI-MS spectra were measured with a Bruker HCT ultra PTM Discovery system (Proteinex fc, UltiMate 3000). Elemental analyses (C, H, and N) were performed on a Perkin-Elmer 2400 Series II CHNS/O analyzer, and the results were within ±0.4% of the calculated values.

Preparation of Arylcarbonyl Chlorides (14–24). Arylcarboxylic acids (**3–13**) were suspended in dry toluene (150 mL) at 20 ± 2 °C. Oxalyl chloride (2.2 equiv) was added dropwise. The reaction mixtures were stirred for 30 min at 20 ± 2 °C and then DMF (2 drops) was added. The mixtures were stirred for 6 h and then evaporated to dryness. The residues were washed with petroleum ether and used directly in the next step.

Preparation of Carboxamides (25–35, 55). Into solutions of **14–24**, **54** (5.1 mmol) in 200 mL of dry toluene were added triethylamine (4 mL) and 2-amino-4,5-methylenedioxy acetophenone (**2**) (5 mmol). The mixtures were stirred at 20 ± 2 °C for 24 h and then evaporated. The residues were washed with acetone and EtOH and then recrystallized from acetone or EtOH to form the pure carboxamides.

N-(6-Acetyl-1,3-benzodioxol-5-yl)-1-benzothiophene-3-carboxamide (25). Obtained as a pale-yellow solid from 2-amino-4,5-methylenedioxyacetophenone (**2**) and benzo[*b*]thiophene-3-carbonyl chloride (**14**); mp 213–214 °C; MS (ESI) 340 *m/z* [M + H]⁺. ¹H NMR (400 MHz, DMSO-*d*₆, δ): 2.63 (3H, s), 6.18 (2H, s), 7.43–7.53 (2H, m), 7.66 (1H, s), 8.10 (1H, dd, *J* = 1.2, 7.2 Hz), 8.30 (1H, s), 8.47 (1H, dd, *J* = 1.2, 7.2 Hz), 8.53 (1H, s), 12.75 (1H, s). IR (KBr): 1638, 1668 (C=O) cm⁻¹.

N-(6-Acetyl-1,3-benzodioxol-5-yl)-1-benzothiophene-2-carboxamide (26). Obtained as a grayish-white solid from **2** and benzo[*b*]thiophene-2-carbonyl chloride (**15**); mp 233–235 °C; MS (ESI) 340 *m/z* [M + H]⁺. ¹H NMR (400 MHz, DMSO-*d*₆, δ): 2.69 (3H, s), 6.19 (2H, s), 7.46–7.57 (2H, m), 7.71 (1H, s), 8.09–8.13 (2H, m), 8.14 (1H, s), 8.25 (1H, s), 13.11 (1H, s). IR (KBr): 1640, 1655 (C=O) cm⁻¹.

N-(6-Acetyl-1,3-benzodioxol-5-yl)-1-benzofuran-3-carboxamide (27). Obtained as a pale-yellow solid from **2** and benzo[*b*]furan-3-carbonyl chloride (**16**); mp 144–145 °C; MS (ESI) 324 *m/z* [M + H]⁺. ¹H NMR (400 MHz, DMSO-*d*₆, δ): 2.63 (3H, s), 6.19 (2H, s), 7.41–7.50 (2H, m), 7.68 (1H, s), 7.75 (1H, dd, *J* = 1.6, 6.8 Hz), 8.15 (1H, dd, *J* = 2.0, 8.8 Hz), 8.27 (1H, s), 8.71 (1H, s), 12.63 (1H, s). IR (KBr): 1635, 1677 (C=O) cm⁻¹.

N-(6-Acetyl-1,3-benzodioxol-5-yl)-1-benzofuran-2-carboxamide (28). Obtained as a grayish-white solid from **2** and benzo[*b*]thiophene-2-carbonyl chloride (**17**); mp 184–185 °C; MS (ESI) 324 *m/z* [M + H]⁺. ¹H NMR (400 MHz, DMSO-*d*₆, δ): 2.69 (3H, s), 6.20 (2H, s), 7.40 (1H, t, *J* = 7.6 Hz), 7.55 (1H, td, *J* = 1.2, 7.8 Hz), 7.72–7.78 (3H, m), 7.84 (1H, d, *J* = 8.0 Hz), 8.36 (1H, s), 13.21 (1H, s). IR (KBr): 1667, 1682 (C=O) cm⁻¹.

N-(6-Acetyl-1,3-benzodioxol-5-yl)naphthalene-1-carboxamide (29). Obtained as a grayish-white solid from **2** and naphthalene-1-carbonyl chloride (**18**); mp 143–144 °C; MS (ESI) 334 *m/z* [M + H]⁺. ¹H NMR (400 MHz, DMSO-*d*₆, δ): 2.59 (3H, s), 6.20 (2H, s), 7.60–7.68 (4H, m), 7.87 (1H, d, *J* = 7.2 Hz), 8.05–8.07 (1H, m), 8.15 (1H, d, *J* = 8.0 Hz), 8.33–8.38 (2H, m), 12.52 (1H, s). IR (KBr): 1647, 1672 (C=O) cm⁻¹.

N-(6-Acetyl-1,3-benzodioxol-5-yl)naphthalene-2-carboxamide (30). Obtained as a pale-yellow solid from **2** and naphthalene-2-carbonyl chloride (**19**); mp 172–173 °C; MS (ESI) 334 *m/z* [M + H]⁺. ¹H NMR (400 MHz, DMSO-*d*₆, δ): 2.68 (3H, s), 6.19 (2H, s), 7.64–7.72 (3H, m), 7.99–8.06 (2H, m), 8.15 (2H, d, *J* = 8.8 Hz), 8.42 (1H, s), 8.58 (1H, s), 13.09 (1H, s). IR (KBr): 1636, 1670 (C=O) cm⁻¹.

N-(6-Acetyl-1,3-benzodioxol-5-yl)quinoline-4-carboxamide (31). Obtained as a grayish-white solid from **2** and quinoline-4-carbonyl chloride (**20**); mp 166–167 °C; MS (ESI) 335 *m/z* [M + H]⁺. ¹H NMR (400 MHz, DMSO-*d*₆, δ): 2.59 (3H, s), 6.21 (2H, s), 7.69 (1H, s), 7.72 (1H, ddd, *J* = 1.2, 7.2 Hz), 7.81 (1H, d, *J* = 4.4 Hz), 7.87 (1H, ddd, *J* = 1.2, 7.4 Hz), 8.15 (1H, d, *J* = 8.4 Hz), 8.24 (1H, s), 8.31 (1H, d, *J* = 8.4 Hz), 9.09 (1H, d, *J* = 4.2 Hz), 12.48 (1H, s). IR (KBr): 1645, 1680 (C=O) cm⁻¹.

N-(6-Acetyl-1,3-benzodioxol-5-yl)quinoline-3-carboxamide (32). Obtained as a pale-yellow solid from **2** and quinoline-3-carbonyl chloride (**21**); mp 215–216 °C; MS (ESI) 335 *m/z* [M + H]⁺. ¹H NMR (400 MHz, DMSO-*d*₆, δ): 2.66 (3H, s), 6.21 (2H, s), 7.73 (1H, s), 7.76 (1H, ddd, *J* = 1.2, 7.4 Hz), 7.94 (1H, ddd, *J* = 1.6, 6.8 Hz), 8.15 (1H, d, *J* = 8.4 Hz), 8.23 (1H, d, *J* = 7.2 Hz), 8.34 (1H, s), 8.92 (1H, s), 9.38 (1H, s), 12.05 (1H, s). IR (KBr): 1639, 1670 (C=O) cm⁻¹.

N-(6-Acetyl-1,3-benzodioxol-5-yl)quinoline-2-carboxamide (33). Obtained as a yellow solid from **2** and quinoline-2-carbonyl chloride (**22**); mp 210–211 °C; MS (ESI) 335 *m/z* [M + H]⁺. ¹H NMR (400 MHz, DMSO-*d*₆, δ): 2.69 (3H, s), 6.21 (2H, s), 7.73 (1H, s), 7.79 (1H, ddd, *J* = 1.2, 7.2 Hz), 7.95 (1H, ddd, *J* = 1.6, 7.6 Hz), 8.15 (1H, d, *J* = 7.2 Hz), 8.22 (1H, d, *J* = 8.0 Hz), 8.28 (1H, d, *J* = 8.8 Hz), 8.55 (1H, s), 8.65–8.68 (1H, d, *J* = 8.8). IR (KBr): 1647, 1670 (C=O) cm⁻¹.

N-(6-Acetyl-1,3-benzodioxol-5-yl)quinoline-5-carboxamide (34). Obtained as a yellow solid from **2** and quinoline-5-carbonyl chloride (**23**); mp 210–211 °C; MS (ESI) 335 *m/z* [M + H]⁺. ¹H NMR (400 MHz, DMSO-*d*₆, δ): 2.61 (3H, s), 6.21 (2H, s), 7.70 (1H, s), 7.82 (1H, dd, *J* = 4.4, 8.8 Hz), 8.03 (1H, t, *J* = 7.8 Hz), 8.11 (1H, d, *J* = 7.2 Hz), 8.30–8.35 (2H, m), 9.03 (1H, d, *J* = 8.4 Hz), 9.13 (1H, d, *J* = 4.4 Hz), 12.59 (1H, s). IR (KBr): 1636, 1672 (C=O) cm⁻¹.

N-(6-Acetyl-1,3-benzodioxol-5-yl)anthracene-1-carboxamide (35). Obtained as a pale-yellow solid from **2** and anthracene-1-carbonyl chloride (**24**); mp 206–207 °C; MS (ESI) 384 *m/z* [M + H]⁺. ¹H NMR (400 MHz, DMSO-*d*₆, δ): 2.59 (3H, s), 6.21 (2H, s), 7.55–7.61 (2H, m), 7.62–7.67 (1H, m), 7.70 (1H, s), 7.90 (1H, d, *J* = 6.8 Hz), 8.13–8.16 (2H, m), 8.32 (1H, d, *J* = 8.4 Hz), 8.46 (1H, s), 8.73 (1H, s), 9.03 (1H, s), 12.60 (1H, s). IR (KBr): 1643, 1674 (C=O) cm⁻¹.

N-(6-Acetyl-1,3-benzodioxol-5-yl)-2-(naphthalen-1-yl)acetamide (55). Obtained as a white solid from **2** and 1-naphthylmethyl chloride (**54**); mp 89–90 °C; MS (ESI) 384 *m/z* [M + H]⁺. ¹H NMR (400 MHz, DMSO-*d*₆, δ): 2.53 (3H, s), 4.21 (2H, s), 6.13 (2H, s), 7.50–7.61 (5H, m), 7.90 (1H, d, *J* = 8.0 Hz), 7.95 (1H, dd, *J* = 2.0, 7.2 Hz), 8.03 (1H, dd, *J* = 1.2, 8.0 Hz), 8.15 (1H, s), 11.89 (1H, s). IR (KBr): 1641, 1684 (C=O) cm⁻¹.

Preparation of 2-Aryl-6,7-methylenedioxyquinolin-4-ones (1, 37–44). Method A. Into a suspension of **25**, **27–34** (2.95 mmol) in *t*-butyl alcohol (100 mL) was added potassium *t*-butoxide (1.66 g, 14.7 mmol). The mixture was refluxed under argon for 12 h, cooled, and poured into a 10% ammonium chloride solution (100 mL). The solid precipitate was collected and washed with EtOH. The crude product was purified by flash chromatography (silica gel, CH₂Cl₂:EtOH 16:1–10:1).

2-(3-Benzo[*b*]thienyl)-6,7-methylenedioxyquinolin-4-one (1). Yield 35% from **25** as a white solid; mp > 330 °C; MS (ESI)

322 m/z [M + H]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.15 (2H, s), 6.19 (1H, s), 7.11 (1H, s), 7.43 (1H, s), 7.47–7.54 (2H, m), 7.93 (1H, d, *J* = 7.6 Hz), 8.14 (1H, d, *J* = 7.6 Hz), 8.24 (1H, s), 11.80 (1H, s). IR (KBr): 1616 (C=O) cm⁻¹. Anal. Calcd for C₁₈H₁₁NO₃S: C, 67.00; H, 3.48; N, 4.25. Found: C, 67.28; H, 3.45; N, 4.36.

2-(3-Benzo[*b*]furyl)-6,7-methylenedioxyquinolin-4-one (37). Yield 17% from **27** as a pale-yellow solid; mp > 315 °C; MS (ESI) m/z 306 [M + H]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.12 (2H, s), 6.49 (1H, s), 7.13 (1H, s), 7.36–7.45 (3H, m), 7.69 (1H, d, *J* = 8.0 Hz), 8.14 (1H, s), 8.52 (1H, s). IR (KBr): 1626 (C=O) cm⁻¹. Anal. Calcd for C₁₈H₁₁NO₄: C, 70.82; H, 3.63; N, 4.59. Found: C, 70.52; H, 3.95; N, 4.21.

2-(2-Benzo[*b*]furyl)-6,7-methylenedioxyquinolin-4-one (38). Yield 28% from **28** as a grayish–white solid; mp > 320 °C; MS (EI, 70 eV) m/z 305 [M]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.19 (2H, s), 6.75 (1H, s), 7.30 (1H, s), 7.35 (1H, t, *J* = 7.6 Hz), 7.40 (1H, s), 7.45 (1H, t, *J* = 7.6 Hz), 7.71–7.79 (3H, m). IR (KBr): 1630 (C=O) cm⁻¹. Anal. Calcd for C₁₈H₁₁NO₄: C, 70.82; H, 3.63; N, 4.59. Found: C, 70.62; H, 3.84; N, 4.24.

2-(1-Naphthalenyl)-6,7-methylenedioxyquinolin-4-one (39). Yield 52% from **29** as a grayish–white solid; mp > 350 °C; MS (ESI) m/z 316 [M + H]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.08 (1H, s), 6.15 (2H, s), 7.03 (1H, s), 7.46 (1H, s), 7.56–7.63 (2H, m), 7.63–7.70 (2H, m), 7.83 (1H, d, *J* = 7.6 Hz), 8.06 (1H, d, *J* = 7.6 Hz), 8.11 (1H, d, *J* = 7.6 Hz), 11.90 (1H, s). IR (KBr): 1653 (C=O) cm⁻¹. Anal. Calcd for C₂₀H₁₃NO₃: C, 76.18; H, 4.16; N, 4.44. Found: C, 75.60; H, 3.94; N, 4.29.

2-(2-Naphthalenyl)-6,7-methylenedioxyquinolin-4-one (40). Yield 48% from **30** as a white solid; mp > 330 °C; MS (ESI) m/z 316 [M + H]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.16 (2H, s), 6.49 (1H, s), 7.25 (1H, s), 7.42 (1H, s), 7.59–7.64 (2H, m), 7.88–7.95 (1H, m), 8.00–8.03 (1H, m), 8.06–8.12 (2H, m), 8.42 (1H, s). IR (KBr): 1616 (C=O) cm⁻¹. Anal. Calcd for C₂₀H₁₃NO₃: C, 76.18; H, 4.16; N, 4.44. Found: C, 76.04; H, 4.28; N, 4.28.

2-(4-Quinolyl)-6,7-methylenedioxyquinolin-4-one (41). Yield 54% from **31** as a pale-yellow solid; mp > 320 °C; MS (ESI) m/z 317 [M + H]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.12–6.29 (3H, m), 7.06 (1H, s), 7.47 (1H, s), 7.65–7.70 (2H, m), 7.86 (1H, t, *J* = 7.6 Hz), 7.93 (1H, d, *J* = 8.4 Hz), 8.15 (1H, d, *J* = 8.0 Hz), 9.03 (1H, s). IR (KBr): 1618 (C=O) cm⁻¹. Anal. Calcd for C₁₉H₁₂N₂O₃: C, 72.15; H, 3.82; N, 8.86. Found: C, 72.35; H, 4.03; N, 8.60.

2-(3-Quinolyl)-6,7-methylenedioxyquinolin-4-one (42). Yield 65% from **32** as a grayish–white solid; mp > 320 °C; MS (ESI) m/z 317 [M + H]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.19 (2H, s), 6.62 (1H, s), 7.23 (1H, s), 7.44 (1H, s), 7.73 (1H, t, *J* = 7.6 Hz), 7.87 (1H, t, *J* = 7.6 Hz), 8.11–8.17 (2H, m), 8.88 (1H, s), 9.38 (1H, s). IR (KBr): 1618 (C=O) cm⁻¹. Anal. Calcd for C₁₉H₁₂N₂O₃: C, 72.15; H, 3.82; N, 8.86. Found: C, 71.86; H, 3.76; N, 8.63.

2-(2-Quinolyl)-6,7-methylenedioxyquinolin-4-one (43). Yield 48% from **33** as a pale-yellow solid. mp 345–347 °C; MS (EI, 70 eV) m/z 316 [M]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.18 (2H, s), 6.98 (1H, s), 7.43 (1H, s), 7.63 (1H, s), 7.73 (1H, t, *J* = 7.6 Hz), 7.91 (1H, t, *J* = 7.6 Hz), 8.11 (1H, d, *J* = 8.0 Hz), 8.27 (1H, d, *J* = 8.4 Hz), 8.32 (1H, d, *J* = 8.8 Hz), 8.60 (1H, d, *J* = 8.8 Hz), 11.85 (1H, s). IR (KBr): 1611 (C=O) cm⁻¹. Anal. Calcd for C₁₉H₁₂N₂O₃: C, 72.15; H, 3.82; N, 8.86. Found: C, 71.76; H, 3.88; N, 8.46.

2-(5-Quinolyl)-6,7-methylenedioxyquinolin-4-one (44). Yield 47% from **34** as a pale-yellow solid; mp > 330 °C; MS (ESI) m/z 317 [M + H]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.03–6.19 (3H, m), 7.03 (1H, s), 7.47 (1H, s), 7.60 (1H, dd, *J* = 4.0, 8.4 Hz), 7.81 (1H, d, *J* = 6.8 Hz), 7.91 (1H, t, *J* = 7.8 Hz), 8.20 (1H, d, *J* = 8.4 Hz), 8.30 (1H, d, *J* = 6.4 Hz), 8.99 (1H, s), 11.91 (1H, s). IR (KBr): 1614 (C=O) cm⁻¹. Anal. Calcd for C₁₉H₁₂N₂O₃: C, 72.15; H, 3.82; N, 8.86. Found: C, 72.08; H, 3.94; N, 8.66.

Preparation of 2-Aryl-6,7-methylenedioxyquinolin-4-ones (1, 36, 39, 45) and 4-Methyl-6,7-methylenedioxy-3-(1-naphthyl)quinolin-2-one (56). Method B. Following the same procedure

described in method A, except for the use of 1,4-dioxane as solvent in place of *t*-butyl alcohol, and the use of NaOH in place of potassium *t*-butoxide. Compounds **1** and **39** were confirmed by comparison of mp and TLC with those of a sample obtained from method A.

2-(3-Benzo[*b*]thienyl)-6,7-methylenedioxyquinolin-4-one (1). Yield 40% from **25** as a pale-yellow solid.

2-(2-Benzo[*b*]thienyl)-6,7-methylenedioxyquinolin-4-one (36). Yield 74% from **26** as a white solid; mp > 350 °C; MS (ESI) m/z 322 [M + H]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.20 (2H, s), 7.21 (1H, s), 7.39–7.45 (3H, m), 7.96–8.16 (3H, m), 11.55 (1H, s). IR (KBr): 1616 (C=O) cm⁻¹. Anal. Calcd for C₁₈H₁₁NO₃S: C, 67.00; H, 3.48; N, 4.25. Found: C, 67.21; H, 3.47; N, 4.33.

2-(1-Naphthalenyl)-6,7-methylenedioxyquinolin-4-one (39). Yield 58% from **29** as a grayish–white solid.

2-(1-Anthracenyl)-6,7-methylenedioxyquinolin-4-one (45). Yield 48% yield from **35** as a yellow solid; mp > 320 °C; MS (ESI) m/z 366 [M + H]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.15 (3H, s), 7.03 (1H, s), 7.43–7.70 (5H, m), 8.08 (2H, t, *J* = 7.6 Hz), 8.25 (1H, d, *J* = 8.4 Hz), 8.47 (1H, s), 8.70 (1H, s), 11.98 (1H, s). IR (KBr): 1632 (C=O) cm⁻¹. Anal. Calcd for C₂₄H₁₅NO₃: C, 78.89; H, 4.14; N, 3.83. Found: C, 78.24; H, 4.40; N, 3.37.

4-Methyl-6,7-methylenedioxy-3-(1-naphthyl)quinolin-2-one (56). Yield 61% from **55** as a white solid; mp > 260 °C; MS (EI, 70 eV) m/z 329 [M]⁺. ¹H NMR (DMSO-*d*₆, δ): 2.02 (3H, s), 6.13 (2H, s), 6.91 (1H, s), 7.29–7.32 (2H, m), 7.39–7.47 (2H, m), 7.52 (1H, ddd, *J* = 1.6, 6.4 Hz), 7.58 (1H, t, *J* = 8.0 Hz), 7.93–8.01 (2H, m), 11.84 (1H, s). IR (KBr): 1632 (C=O) cm⁻¹. Anal. Calcd for C₂₁H₁₅NO₃: C, 76.58; H, 4.59; N, 4.25. Found: C, 75.88; H, 5.07; N, 4.06.

Deacetylation of Compounds 25, 26, 28–32, and 34. Into a suspension of compound (2.95 mol) in dry toluene (150 mL) was added potassium *t*-butoxide (1.66 g, 14.75 mol). The mixture was heated under argon at 90 °C for 72 h, and then concentrated and neutralized with 20% HOAc. The resulting solid precipitate was collected and purified by flash chromatography (silica gel, CH₂Cl₂–EtOH) to afford **46–53**.

N-(1,3-Benzodioxol-5-yl)-1-benzothiophene-3-carboxamide (46). Yield 28% from **25** as a white solid; mp 188–189 °C; MS (EI, 70 eV) m/z 297 [M]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.01 (2H, s), 6.90 (1H, d, *J* = 8.4 Hz), 7.17 (1H, dd, *J* = 2.0, 8.4 Hz), 7.40–7.51 (3H, m), 8.06 (1H, dd, *J* = 1.2, 7.2 Hz), 8.37 (1H, dd, *J* = 0.8, 8.0 Hz), 8.46 (1H, s), 10.29 (1H, s). IR (KBr): 1647 (C=O) cm⁻¹. Anal. Calcd for C₁₆H₁₁NO₃S: C, 64.63; H, 3.73; N, 4.71. Found: C, 64.32; H, 3.94; N, 4.50.

N-(1,3-Benzodioxol-5-yl)-1-benzothiophene-2-carboxamide (47). Yield 36% from **26** as a grayish–white solid; mp 171–173 °C; MS (EI, 70 eV) m/z 297 [M]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.01 (2H, s), 6.91 (1H, d, *J* = 8.4 Hz), 7.15 (1H, dd, *J* = 2.0, 8.4 Hz), 7.38 (1H, d, *J* = 2.4 Hz), 7.41–7.54 (2H, m), 7.96–8.05 (2H, m), 8.28 (1H, s), 10.41 (1H, s). IR (KBr): 1634 (C=O) cm⁻¹. Anal. Calcd for C₁₆H₁₁NO₃S: C, 64.63; H, 3.73; N, 4.71. Found: C, 64.28; H, 3.98; N, 4.46.

N-(1,3-Benzodioxol-5-yl)-1-benzofuran-2-carboxamide (48). Yield 28% from **28** as a yellow solid; mp 156–158 °C; MS (EI, 70 eV) m/z 281 [M]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.03 (2H, s), 6.93 (1H, d, *J* = 8.4 Hz), 7.27 (1H, dd, *J* = 2.0, 8.4 Hz), 7.37 (1H, t, *J* = 7.6 Hz), 7.47 (1H, d, *J* = 2.0 Hz), 7.51 (1H, ddd, *J* = 2.0, 8.4 Hz), 7.71–7.75 (2H, m), 7.82 (1H, d, *J* = 8.0 Hz), 10.49 (1H, s). IR (KBr): 1668 (C=O) cm⁻¹. Anal. Calcd for C₁₆H₁₁NO₄: C, 68.32; H, 3.94; N, 4.98. Found: C, 68.22; H, 4.14; N, 4.22.

N-(1,3-Benzodioxol-5-yl)naphthalene-1-carboxamide (49). Yield 32% from **29** as a white solid; mp 204–206 °C; MS (EI, 70 eV) m/z 291 [M]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.02 (2H, s), 6.91 (1H, d, *J* = 8.4 Hz), 7.22 (1H, dd, *J* = 2.0, 8.4 Hz), 7.51 (1H, d, *J* = 2.0 Hz), 7.56–7.63 (3H, m), 7.72 (1H, d, *J* = 6.4 Hz), 7.99–8.03 (1H, m), 8.06 (1H, d, *J* = 8.4 Hz), 8.16–8.20 (1H, m), 10.46 (1H, s). IR (KBr): 1643 (C=O) cm⁻¹. Anal. Calcd for C₁₈H₁₃NO₃: C, 74.22; H, 4.50; N, 4.81. Found: C, 73.83; H, 4.82; N, 4.26.

N-(1,3-Benzodioxol-5-yl)naphthalene-2-carboxamide (50). Yield 21% yield from **30** as a pale-yellow solid; mp 184–185 °C;

MS (EI, 70 eV) m/z 291 [M]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.02 (2H, s), 6.94 (1H, d, *J* = 8.4 Hz), 7.24 (1H, dd, *J* = 2.0, 8.4 Hz), 7.50 (1H, d, *J* = 2.0 Hz), 7.60–7.68 (2H, m), 7.98–8.15 (4H, m), 8.56 (1H, s), 10.38 (1H, s). IR (KBr): 1636 (C=O) cm⁻¹. Anal. Calcd for C₁₈H₁₃NO₃: C, 74.22; H, 4.50; N, 4.81. Found: C, 74.19; H, 4.62; N, 4.56.

***N*-(1,3-Benzodioxol-5-yl)quinoline-4-carboxamide (51)**. Yield 31% from **31** as a white solid; mp 167–168 °C; MS (EI, 70 eV) m/z 292 [M]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.02 (2H, s), 6.93 (1H, d, *J* = 8.4 Hz), 7.16 (1H, dd, *J* = 1.6, 8.4 Hz), 7.43 (1H, d, *J* = 2.0 Hz), 7.66–7.72 (2H, m), 7.84 (1H, t, *J* = 8.0 Hz), 8.11 (2H, d, *J* = 8.8 Hz), 9.01 (1H, s), 10.65 (1H, s). IR (KBr): 1636 (C=O) cm⁻¹. Anal. Calcd for C₁₇H₁₂N₂O₃: C, 69.86; H, 4.14; N, 9.58. Found: C, 69.65; H, 4.32; N, 9.34.

***N*-(1,3-Benzodioxol-5-yl)quinoline-3-carboxamide (52)**. Yield 23% from **32** as a yellow solid; mp 254–256 °C; MS (EI, 70 eV) m/z 292 [M]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.05 (2H, s), 6.95 (1H, d, *J* = 8.4 Hz), 7.23 (1H, dd, *J* = 2.0, 8.4 Hz), 7.49 (1H, d, *J* = 2.0 Hz), 7.74 (1H, ddd, *J* = 1.2, 8.0 Hz), 7.90 (1H, ddd, *J* = 1.6, 8.4 Hz), 8.10–8.18 (2H, m), 8.92 (1H, s), 9.33 (1H, s), 10.56 (1H, s). IR (KBr): 1659 (C=O) cm⁻¹. Anal. Calcd for C₁₇H₁₂N₂O₃: C, 69.86; H, 4.14; N, 9.58. Found: C, 69.73; H, 4.35; N, 9.42.

***N*-(1,3-Benzodioxol-5-yl)quinoline-5-carboxamide (53)**. Yield 20% from **34** as a white solid; mp 202–203 °C; MS (EI, 70 eV) m/z 292 [M]⁺. ¹H NMR (DMSO-*d*₆, δ): 6.02 (2H, s), 6.95 (1H, d, *J* = 8.4 Hz), 7.21 (1H, dd, *J* = 2.0, 8.0 Hz), 7.50 (1H, d, *J* = 2.0 Hz), 7.63 (1H, dd, *J* = 4.0, 8.4 Hz), 7.83–7.90 (2H, m), 8.16–8.20 (1H, m), 8.63 (1H, d, *J* = 8.0 Hz), 8.98 (1H, dd, *J* = 1.6, 4.0 Hz), 10.58 (1H, s). IR (KBr): 1643 (C=O) cm⁻¹. Anal. Calcd for C₁₇H₁₂N₂O₃: C, 69.86; H, 4.14; N, 9.58. Found: C, 69.68; H, 4.33; N, 9.52.

Biological Evaluations. MTT (3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) Assays.^{25,26} HL-60, HCT-116, and Hep 3B cells were treated with tested compounds for the indicated periods. After treatment, cells were washed once with PBS and incubated with MTT (Sigma, St. Louis, MO) for 2 h. The formazan precipitate was dissolved in 150 μL of DMSO, and the absorbance was measured with an ELISA reader at 570 nm.

SRB Assays.²⁷ A549, KB, KB-VIN, and DU145 cells were treated with tested compounds for the indicated periods. After additional incubation with DMSO or compounds, cells were fixed with 10% TCA, and SRB at 0.4% (w/v) in 1% acetic acid was added to stain cells. Unbound SRB was washed out with 1% acetic acid, and SRB bound to the cells was solubilized with 10 mM Trizma base. Absorbance was read at 515 nm.

Tubulin Assays.^{25,26} Tubulin assembly was measured by turbidimetry as described in detail previously.²⁸ Assay mixtures contained 1.0 mg/mL (10 μM) tubulin and varying drug concentrations and were preincubated 15 min at 30 °C in the absence of GTP. The samples were placed on ice, and 0.4 mM GTP was added. Reaction mixtures were transferred to cuvettes held at 0 °C, and turbidity development was followed for 20 min at 30 °C after a rapid temperature jump. Drug concentrations that inhibited increase in turbidity by 50% relative to a control sample were determined.

Inhibition of the binding of [³H]colchicine to tubulin was measured as described in detail previously.²⁹ Incubation of 1.0 μM tubulin with 5.0 μM [³H]colchicine and either 1.0 or 5.0 μM inhibitor was for 10 min at 37 °C. At this time point, approximately 40–60% of maximum colchicine binding occurs.

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